

Flexing FLEX for Suborbital Studies: Biological Imaging & More

Jordan Callaham¹, Sarah Swanson⁴, Simon Gilroy⁴, Anna-Lisa Paul^{1,2} and Robert Ferl^{1,3}

1. Department of Horticultural Sciences, University of Florida, Gainesville, FL USA 2. ICBR, University of Florida, Gainesville, FL USA 3. Office of Research, University of Florida, Gainesville, FL USA

4. Department of Botany, University of Wisconsin, Madison, WI USA

Abstract

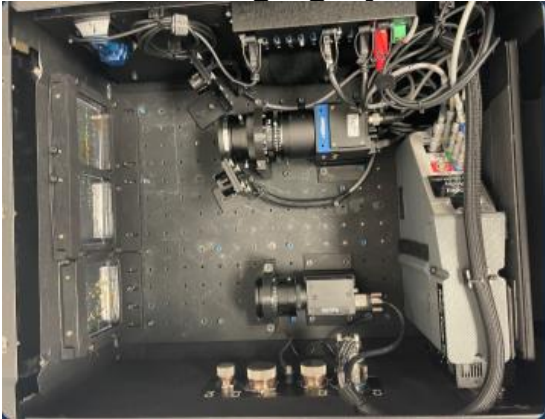
Space launch and space travel's changes in gravitational profiles are unique and cannot currently be found or simulated in Earth-based environments. Suborbital flights, allow experiments to undergo the physical demands and gravitational changes that a rocket launch, transition to a weightless environment, and landing of a spacecraft encompasses. This express, but full simulation, of launch, microgravity, reentry and landing allow for the study of plants outside their evolutionary developed environment. This study addresses both hardware composition of a non-tended launch environment and the biological specimens utilized to study the response to the environment.

biological signals occur during various phases of the flight. Calcium signaling has been shown to be responsive to light, gravity, and other biotic or abiotic stresses when applied to plants. The calcium signaling response, ubiquitous in plant tissues, allows to plants to respond to stimuli intercellularly, both at a tissue specific and whole plant level.

Conclusions

FLEX fluorescence imaging results indicate that plants are responsive to the suborbital spaceflight environment through changes in calcium signaling during the three distinct phases of the flight (launch, zero-g, and re-entry/landing). Launch hyper-g's show a decrease in overall GCaMP3 expression, followed by increases both generally and with tissue specific localization during the zero-g and re-entry portions of flight.

FLEX Imaging System

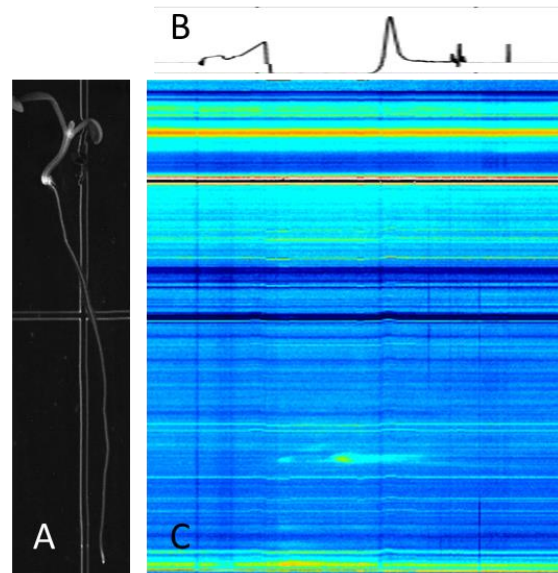


Hardware Development

Ongoing and continuous updates and adaptation to the FLEX hardware have been made in support of the P11 and P12 Blue Origin payload flights. Adaptions by way of removing a thermal imager and the addition of a second monochrome camera for GFP imaging. Both monochrome cameras are currently fitted with 520bp20 filters for optimal imaging. Additionally, a second LED pod for illumination was added for the secondary camera. A Surface 8 computer and MSR device are internal support hardware, both flown on previous flights.

Biological Specimen

Arabidopsis thaliana, modified with the GCaMP3 construct to express Green Fluorescent Protein (GFP) induced by calcium signaling, was used to demonstrate that rapid (~1 second resolution)



Data generated from FLEX on P11 flight for a single GCaMP WT plant during the flight. **A.** Cropped image of the representative plant. **B.** Gravitational data of the P11 flight. **C.** X – Average GFP expression intensity (by color) of the plant (correlated to the row average of plant expression intensity.) Y – Each column represents one second of flight (corresponding to G-data in B.)

This work is supported by NASA Flight Opportunities grant 80NSSC20K0113 and NASA Biological and Physical Sciences grant 80NSSC19K1500 to A-L. Paul and R. Ferl.